

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant(s) : Wei et al.  
U.S. Serial No. : 10/650,365  
Confirmation No. : 7677  
Filed : August 28, 2003  
Examiner : Jegatheesan Seharaseyon  
Art Unit : 1647  
For : RECOMBINANT SUPER-COMPOUND INTERFERON

Law Offices of Albert Wai-Kit Chan, LLC  
World Plaza, Suite 604  
141-07 20<sup>th</sup> Avenue  
Whitestone, New York 11357

July 3, 2007

Commissioner for Patents  
P.O. Box 1450  
Alexandria, VA 22313-1450

Dear Sir/Madam:

SUPPLEMENTAL RESPONSE TO AUGUST 23, 2005 OFFICE ACTION

This Amendment is being submitted as a Supplemental Response to the August 23, 2005 Office Action which was issued by the United States Patent and Trademark Office (USPTO) in connection with the above-identified application.

Priority

The Examiner acknowledged Applicants' claim for foreign priority based on an application filed in China on February 28, 2001. However, the Examiner also noted that Applicants did not file a

Applicant(s) : Wei et al.  
U.S. Serial No.: 10/650,365  
Filed : August 28, 2003  
Page : 2

translated copy of the Application No. CHINA 01104367.9. Therefore the priority is set forth as the filing date of the instant application. Consequently, Applicants respectfully submit **Exhibit 1** (16 pages), which is the European counterpart of Applicant's International application referring to the same invention. It serves as the English translation of PCT/CN02/00128 and claims priority to Chinese Patent CN 01104367.9, filed on February 28, 2001.

If a telephone interview would be of assistance in advancing the prosecution of the subject application, Applicants' undersigned attorney invites the Examiner to telephone him at the number provided below. If any additional fee is required, authorization is hereby given to charge the amount of any such fee to Deposit Account No. 50-1891.

Respectfully submitted,

Albert Wai Kit Chan

Albert Wai-Kit Chan  
Registration No. 36,479  
Attorney for Applicant(s)  
Law Offices of  
Albert Wai-Kit Chan, PLLC  
World Plaza, Suite 604  
141-07 20<sup>th</sup> Avenue  
Whitestone, New York 11357  
Tel: (718) 799-1000  
Fax: (718) 357-8615  
E-mail: chank@kitchanlaw.com

# **EXHIBIT 1**



## Description

## FIELD OF THE INVENTION

[0001] This invention is related to a recombinant super-compound interferon (rSIFN-co) with changed dimensional structure. The characteristic of rSIFN-co in this invention is that it cannot only inhibit DNA (deoxyribonucleic acid) duplication of the hepatitis B virus but also the secretion of HBsAg and HBeAg.

## BACKGROUND OF THE INVENTION

[0002] rSIFN-co is a new interferon molecule constructed with the most popular conservative amino acid found in natural human  $\alpha$ -IFN subtypes using genetic engineering methods. United States Patent Nos. 4,895,823 and 4,897,471 have described it. rSIFN-co had been proved to have broad-spectrum IFN activity and virus- and tumor-inhibition and natural killer cell activity. United States Patent No. 5,372,808 by Amgen, Inc. addresses treatment of rSIFN-co. Chinese Patent No. 97193605.8 by Amgen, Inc. addresses re-treatment of rSIFN-co on hepatitis C. Chinese Patent No. 96114663.5 by Shenzhen Jusheng Bio-engineering Ltd. addresses treatment of rSIFN-co on hepatitis B and hepatitis C.

[0003] The United States Food and Medicine Administration (FDA) authorized Amgen Ltd. to produce rSIFN-co with *E. coli* for clinical hepatitis C treatment at the end of 1997.

[0004] Hepatitis B patients can be identified when detecting HBsAg and the HBeAg.  $\alpha$ -IFN is commonly used in clinics to treat hepatitis B. IFN binds superficial cell membrane receptors, inhibiting DNA and RNA (ribonucleic acid) duplication, including inducing some enzymes to prevent duplication of the virus in hepatitis-infected cells. All IFNs can inhibit only the DNA duplication of viruses, not the e and s antigen.

## DETAILED DESCRIPTION OF THE INVENTION

## Invention Component

[0005] It was surprising to find that rSIFN-co, the dimensional structure of which has been changed, is not only a preparation to inhibit the DNA duplication of hepatitis B, but to inhibit the secretion of HBsAg and HBeAg.

[0006] The objective of this invention is to offer a preparation of rSIFN-co to inhibit the DNA duplication of hepatitis B viruses and the secretion of HBsAg and HBeAg of hepatitis B and decrease them to normal levels.

[0007] *Results of this invention:* The production of rSIFN-co with recombinant techniques. On the condition of fixed amino acid sequence, the IFN DNA was redesigned according to the *E. coli* codon usage and then the rSIFN-co gene was artificially synthesized.

[0008] rSIFN-co cDNA was cloned into the high-expression vector of *E. coli* by DNA recombinant techniques, and a high expression of rSIFN-co was gained by using of induce/activate-mechanism of L-arabinose to activate the transcription of  $P_{BAD}$  promoter.

[0009] Compared with usual thermo-induction, pH induction and IPTG induction systems of genetic engineering, arabinose induction/activation system has some advantages: (1) Common systems relieve promoter function by creating a "derepression" pattern. Promoters then induce downstream gene expression. So temperature and pH change and the addition of IPTG cannot activate promoters directly. In the system disclosed herein, L-arabinose not only deactivates and represses but also activates the transcription of  $P_{BAD}$  promoter which induce a high expression of rSIFN-co. Therefore, the arabinose induction/activation system is a more effective expression system. (2) The relation between Exogenous and L-arabinose dosage is linearly. This means the concentration of arabinose can be changed to adjust the expression level of the exogenous gene. Therefore, it is easier to control the exogenous gene expression level in *E. coli* by arabinose than by changing temperature and pH value. This characteristic is significant for the formation of inclusion bodies. (3) L-arabinose is resourceful cheap and safe, which, on the contrary, are the disadvantages of other inducers such as IPTG.

[0010] This invention creates an effective and resistant rSIFN-co-expressing *E. coli* engineering strain with an L-arabinose induction/activation system. The strain is cultivated and fermented under suitable conditions to harvest the bacterial bodies. Inclusion bodies are then purified after destroying bacteria and washing repeatedly. The end result, mass of high-purity, dimensional-structure-changed rSIFN-co protein for this invention and for clinical treatment, was gained from denaturation and renaturation of inclusion bodies and a series of purification steps.

[0011] The following are some rSIFN-co preparations: tablets, capsules, oral liquids, pastes, injections, sprays, suppositories, and solutions. Injections are recommended. It is common to subcutaneously inject or vein-inject the medicine. The medicine carrier could be any acceptance medicine carrier, including carbohydrate, cellulose, adhesive, collapse, emollient, filling, add-dissolve agent, amorization, preservative, add-thick agent, matching, etc.

## DETAILED DESCRIPTION OF THE FIGURE

[0012] Figure 1. DNA coding sequence and deduced amino acid sequence of rSIFN-co

## EXPERIMENTAL DETAILS

## Embodiment experience:

[0013] The invention disclosed herein also experimentally verifies that the dimensional-structure-changed rSIFN-co can inhibit HBV-DNA duplication and secretion of HBsAg and HBeAg.

## Materials

[0014] Solvent and Dispensing Method: Add 1ml saline into each vial, dissolve, and mix with MEM culture medium at different concentrations. Mix on the spot.

[0015] Control drugs: IFN- $\alpha$ 2b (Intron A) as lyophilized, purchased from Schering Plough.  $3 \times 10^6$ U each, mix to  $3 \times 10^6$ U/ml with culture medium; INFERGEN (liquid solution), purchased from Amgen, 9 $\mu$ g, 0.3ml each, equal to  $9 \times 10^6$ U, and mix with  $9 \times 10^6$ U/ml culture medium preserve at 4°C; 2.2.15 cell; 2.2.15 cell line of hepatoma (Hep G2) cloned and transfected by HBV DNA, constructed by Mount Sinai Medical Center.

[0016] Reagent: MEM powder, Gibco American Ltd, cattle fetal blood serum, HycloneLab American Ltd. G-418 (Geneticin); MEM dispensing, Gibco American Ltd.; L-Glutamyl, Imported and packaged by JING KE Chemical Ltd.; HBsAg and HBeAg solid-phase radioimmunoassay box, Northward Reagent Institute of Chinese Isotope Ltd.; Biogranectina, Northern China Medicine; And Lipofectin, Gibco American Ltd.

[0017] Experimental goods and equipment: culture bottle, Denmark Tuncion™; 24-well and 96-well culture board, Corning American Ltd.; Carbon Dioxide hatching box, Shel-Lab American Ltd.; MEM culture medium 100ml; 10% cattle fetal blood serum, 3% Glutamy1%, G418 980 $\mu$ g/ml, biogranectina 50U/ml.

## Method:

[0018] 2.2.15 cell culture: Added 0.25% pancreatic enzyme into culture box with full of 2.2.15 cell, digest at 37°C for 3 minutes, and add culture medium to stop digest and disturb it to disperse the cells, reproduce with ratio of 1:3. They will reach full growth in 10 days.

[0019] Medicine toxicity test: In this test, set groups of different medicine concentrations and a control group in which cell is not acted on with medicine. Digest cell, and dispense to a 100,000 cell/ml solution. Inoculate to 96-well culture board, 200 $\mu$ l each well, culture at 37°C for 24h with 5% CO<sub>2</sub>. Test when simple cell layer grows.

[0020] Dispense rSIFN-co to  $1.8 \times 10^7$ U/ml solution then prepare a series of solutions diluted at two-fold gradients. Add into 96-well culture board, 3 wells per concentration. Change the solution every 4 days. Test cytopathic effect by microscope after 8 days. Fully destroy as 4, 75% as 3, 50% as 2, 25% as 1, zero as 0. Calculate average cell lesion and inhibition rate of different concentrations. Calculate TC50 and TCO according to the Reed Muench method.

$$TC50 = \text{Antilog} \left( B + \frac{50-B}{A-B} \times C \right)$$

A=log >50 % medicine concentration, B=log <50 % medicine concentration, C=log dilution power

[0021] Inhibition test for HBsAg and HBeAg: Separate into positive and negative HBsAg and HBeAg contrast groups, cell contrast group and medicine concentration groups. Inoculate 700,000 cells/ml of 2.2.15 cell into 6-well culture board, 3 ml each well, culture at 37°C for 24h with 5% CO<sub>2</sub>, then prepare 6 gradually diluted solutions with 3-fold as the grade (Prepare 5 solutions, each with a different protein concentration. The concentration of Solution 2 is 3 times lower than that of Solution 1, the concentration of Solution 3 is 3 times lower than that of Solution 2, etc.)  $4.5 \times 10^6$ U/ml,  $1.5 \times 10^6$ U/ml,  $0.5 \times 10^6$ U/ml,  $0.17 \times 10^6$ U/ml, and  $0.056 \times 10^6$ U/ml, 1 well per concentration, culture at 37°C for 24h with 5% CO<sub>2</sub>. Change solutions every 4 days using the same solution. Collect all culture medium on the 8th day. Preserve at -20°C Repeat test 3 times to estimate HBsAg and HBeAg with solid-phase radioimmunoassay box (Northward Reagent Institute of Chinese Isotope Ltd.). Estimate cpm value of each well with a  $\gamma$ -counting machine.

[0022] Medicinal effects calculation: Calculate cpm mean value of contrast groups and different-concentration groups and their standard deviation, P/N value such as inhibition rate, IC50 and SI.

$$1) \text{ Antigen inhibition rate (\%)} = \frac{A-B}{A} \times 100$$

- [0023] A = cpm of control group; B = cpm of test group;  
2) Counting the half-efficiency concentration of the medicine

$$\text{Antigen inhibition IC50} = \text{Antilog} \left( B + \frac{50-B}{A-B} \times C \right)$$

A=log>50% medicine concentration, B=log<50 % medicine concentration, C=log dilution power

- 3) SI of interspace-conformation changed rSIFN-co effect on HBsAg and HBeAg in 2.2.15 cell culture:

$$SI = \frac{TC50}{IC50}$$

- 4) Estimate the differences in cpm of each dilution degree from the control group using student t test

[0024] Southern blot: (1) HBV-DNA extract in 2.2.15 cell: Culture cell 8 days. Exsuction culture medium (Separate cells from culture medium by means of draining the culture medium.). Add lysis buffer to break cells, then extract 2 times with a mixture of phenol, chloroform and isoamyl alcohol (1:1:1), 10,000g centrifuge. Collect the supernatant adding anhydrous alcohol to deposit nucleic acid. Vacuum draw, redissolve into 20μl TE buffer. (2) Electrophoresis: Add 6X DNA loading buffer, electrophoresis on 1.5% agarose gel, IV/cm, at fixed pressure for 14-18h. (3) Denaturation and hybridization: respectively dip gel into HCl, denaturation buffer and neutralization buffer. (4) Transmembrane: Make an orderly transfer of DNA to Hybond-N membrane. Bake, hybridize and expose with dot blot hybridization. Scan and analyze relative density with gel-pro software. Calculate inhibition rate and IC50.

#### Results

[0025] Results from Tables 1, 2 and 3 show: After maximum innocuous concentration exponent culturing for 8 days with 2.2.15 cell, the maxima is  $9.0 \pm 0 \times 10^6$  IU/ml and the average inhibition rate of maximum innocuous concentration rSIFN-co to HBsAg is  $46.0 \pm 5.25\%$  ( $P < 0.001$ ), IC50 is  $4.54 \pm 1.32 \times 10^6$  IU/ml, SI is 3.96; rate to HBeAg is  $44.8 \pm 6.6\%$ , IC50 is  $6.49 \pm 0.42 \times 10^6$  IU/ml, SI is 2.77. This shows that rSIFN-co can significantly inhibit the activity of HBeAg and HBsAg, but that the IFN of the contrast group and INTERFERON cannot. It has also been proved in clinic that rSIFN-co can decrease HBeAg and HBsAg or return them to normal levels.

[0026] The following are some examples for the preparation of rSIFN-co:

#### Example 1: Preparation of lyophilized injection

##### [0027]

- |  |                      |        |
|--|----------------------|--------|
| a) rSIFN-co  | $3 \times 10^6$ IU   |        |
| b) citric acid   | 0.2 mg               |        |
| c) dibasic sodium phosphate                              |                      | 2.5 mg |
| d) NaCl  | 4.0 mg               |        |
| e) dextran   | 20 mg                |        |
| f) Polyoxyethelene anhydrosorbitol monoelaso-acids ester | 0.1 ml               |        |
| g) inject water  | to a level of 1.0 ml |        |

[0028] Preparation technique: Weigh materials according to recipe. Dissolve with sterile and pyrogen-free water. Filter through 0.22μm membrane to de-bacterialize, preserve at 6-10°C. Fill in vials after affirming it is sterile and pyrogen-free. Add 1.0 ml solution to each bottle, and lyophilize in freeze dryer.

#### Example 2: Preparation of liquid injection

##### [0029]

- |  |                    |        |
|--|--------------------|--------|
| a) rSIFN-co  | $3 \times 10^6$ IU |        |
| b) citric acid   | 0.2 mg             |        |
| c) dibasic sodium phosphate                              |                    | 2.5 mg |
| d) NaCl  | 4.0 mg             |        |
| e) dextran   | 20 mg              |        |
| f) Polyoxyethelene anhydrosorbitol monoelaso-acids ester | 0.1 ml             |        |

EP 1 371 373 A1

g) inject water to a level of 1.0 ml

[0030] Preparation technique: Weigh materials according to recipe. Dissolve with sterile and pyrogen-free water. Filter through 0.22µm membrane to de-bacterialize, preserve at 8-10°C. Fill in airtight vial after affirming it is sterile and non-pyrogen at 1.0 ml per vial. Store and product at 2-10°C, and protect from light.

10

15

20

25

30

35

40

45

50

55



Table 1. Results of inhibition rate of rSTW-co to HBsAg and HBeAg  
First batch: (rSTW-co)

Inhibition effect to HsAg											
Concentration ( $\times 10^4$ /ml)		Inhibition rate						Average inhibition rate		1- Accumulation	Accumulated inhibition rate
		First well	Second well	Third well	First well	Second well	Third well	First well	Second well		
900	9036	8976	10476	0.436227	0.43935	0.345659	0.407079	0.945909	0.592921	0.614693546	
300	9616	12082	10098	0.3593754	0.245347	0.369269	0.337997	0.5388299	1.254924	0.300392321	
100	9822	16002	12800	0.386508	0.0005	0.2005	0.195836	0.200833	2.054088	0.08867118	
33.33333	15770	15305	16824	0.014991	0	0	0.004997	0.0049959	3.054091	0.001633453	
11.11111	19372	22270	18934	0	0	0	0	0	4.054091	0	
Control	Cell	16010	Blank	0	0	0	0	0	IC50	602.74446016	
Inhibition effect to HBeAg											
Concentration ( $\times 10^4$ /ml)		Inhibition rate						Average inhibition rate		1- Accumulation	Accumulated inhibition rate
		First well	Second well	Third well	First well	Second well	Third well	First well	Second well		
900	7706	7240	7114	0.342155	0.381936	0.392493	0.372261	0.922288	0.627719	0.595006626	
300	8856	7778	9476	0.2439816	0.336008	0.191053	0.257014	0.549972	1.370724	0.286349218	
100	10818	10720	10330	0.07649	0.084856	0.118149	0.093165	0.292983	2.27756	0.133977019	
33.33333	10744	11114	10570	0.002807	0.051221	0.097661	0.07723	0.1598179	3.20033	0.008767408	
11.11111	10672	9352	10810	0.008953	0.201639	0.077173	0.122588	0.122588	4.077742	0.02918541	
Control	Cell	13714	Blank	0	0	0	0	0	IC50	641.7736749	

Second batch: (BRIFW-co)

Inhibition effect to HBeAg									
Concentration ( $\times 10^4$ IU/ml)	First well	Second well	Third well	Inhibition rate			Average inhibition rate	Accumulation	1- Accumulation
				First well	Second well	Third well			
900	7818	8518	9350	0.554378	0.514592	0.467084	0.512068	1.371181	0.487992
300	10344	10628	9160	0.4103867	0.394209	0.477884	0.427497	0.859791	1.850196
100	12296	10228	13262	0.299134	0.18901	0.244072	0.244072	0.4316522	1.916423
33.33333	15364	17414	16188	0.124259	0.00741	0.77291	0.069653	0.1876045	2.74577
11.11111	17386	13632	15406	0.009006	0.222982	0.123865	0.117953	0.117953	3.628819
Control	Cell	16962	Blank	0			Dilution	3	IC50
									365.9357645
Inhibition effect to HBeAg									
Concentration ( $\times 10^4$ IU/ml)	First well	Second well	Third well	Inhibition rate			Average inhibition rate	Accumulation	1- Accumulation
				First well	Second well	Third well			
900	5784	6198	5792	0.498265	0.462353	0.497571	0.486063	0.892477	0.519397
300	7150	8534	8318	0.379771	0.259715	0.278452	0.30598	0.4074138	1.207957
100	9830	11312	10210	0.147284	0.027412	0.11433	0.096345	0.101434	2.111612
33.33333	13942	12368	13478	0	0	0	0	0.0050891	3.111612
11.11111	12418	11634	11352	0	0	0.01267	0.005089	0.005089	4.106523
Control	Cell		Blank	0			Dilution	3	IC50
									611.0919568



Table 2: Results of inhibition rate of Intrun A (IFN- $\alpha$ 2b) to HbsAg and HBeAg

Concentration ( $\times 10^4$ IU/ml)	Inhibition effect to HbsAg						1- Accumulation	Accumulated inhibition rate	
	Inhibition rate			Average inhibition rate	Accumulation				
	First well	Second well	Third well						
300	14918	11724	9550	0	0.029711	0.376229	0.068747	0.93253	0.068746724
100	14868	16890	15182	0	0	0	0	1.93253	0
33.33333	16760	21736	16400	0	0	0	0	2.93253	0
11.11111	20854	15042	16168	0	0	0	0	3.93253	0
3.703704	12083	12063	12063	0	0	0	0	4.93253	0
Control	Cell	17544	Blank	0	0	0	0	IC50	FALSE

Concentration ( $\times 10^4$ IU/ml)	Inhibition effect to HBeAg						1- Accumulation	Accumulated inhibition rate	
	Inhibition rate			Average inhibition rate	Accumulation				
	First well	Second well	Third well						
300	9226	9196	9658	0.152489	0.247108	0.521054	0.189295	0.8292	0.185857736
100	10946	10340	10828	0	0.050156	0.364272	0.018495	0.0184947	0.010110817
33.33333	12250	12390	13934	0	0	0	0	2.810705	0
11.11111	12634	12342	12900	0	0	0	0	3.810705	0
3.703704	10886	10886	10886	0	0	0	0	4.810705	0
Control	Cell	10886	Blank	0	0	0	0	IC50	FALSE

Table 3: Results of inhibition rate of Interferon to HBsAg and HBeAg  
First batch: (Interferon)

Concentration ( $\times 10^4$ IU/ml)	Inhibition effect to HBsAg						1- Accumulation	Accumulation rate	Accumulated inhibition rate
	First well	Second well	Third well	First well	Second well	Third well			
900	14172	12156	17306	0.091655	0.220869	0	0.104175	0.306157	0.254710374
300	12350	12288	16252	0.1417767	0.212409	0	0.118962	0.2019827	0.102024519
100	14164	18934	14194	0.079349	0	0.090245	0.065331	0.082921	0.029916678
33.33333	35722	16034	16340	0	0	0	0	0.0273897	0.007106592
11.11111	17594	17652	14320	0	0	0.082169	0.02739	0.02739	0.005802377
Control	Cell	15602	Blank	0	0	0.082169	0.02739	0.02739	0.005802377
Dilution 3 IC50 FALSE									
Concentration ( $\times 10^4$ IU/ml)	Inhibition effect to HBeAg						1- Accumulation	Accumulation rate	Accumulated inhibition rate
	First well	Second well	Third well	First well	Second well	Third well			
900	12060	11692	12234	0	0.01275	0	0.00425	0.025163	0.024667111
300	12840	11484	12150	0	0.030313	0	0.010104	0.0209125	0.010422073
100	12894	14096	15088	0	0	0	0	0.010808	0.003606955
33.33233	15032	12928	13020	0	0	0	0	0.0108081	0.002704416
11.11111	11794	11584	11508	0.004137	0	0	0	0.010808	0.002167838
Control	Cell	11843	Blank	0	0.028287	0.028287	0.010808	0.010808	0.002167838
Dilution 3 IC50 FALSE									

Second batch: (Infergen)

Concentration ( $\times 10^4$ IU/ml)	Inhibition effect to HBeAg						Average inhibition rate	Accumulation	1- Accumulation	Accumulated inhibition rate
	First well	Second well	Third well	First well	Second well	Third well				
900	6278	6376	6408	0.200031	0.187864	0.183486	0.190367	0.274635	0.809633	0.253290505
300	7699	9092	6394	0.0198777	0	0.18527	0.068283	0.0842678	1.74125	0.046151005
100	8960	7474	8190	0	0.047655	0	0.015685	0.015685	2.725365	0.003794856
33.3333	8530	8144	9682	0	0	0	0	0	3.725365	0
11.1111	7848	7848	7848	0	0	0	0	0	4.725365	0
Control	Cell	7848		Blank	0		Dilution	3	IC50	FALSE

  

Concentration ( $\times 10^4$ IU/ml)	Inhibition effect to HsAg						Average inhibition rate	Accumulation	1- Accumulation	Accumulated inhibition rate
	First well	Second well	Third well	First well	Second well	Third well				
900	12364	12268	12274	0.036171	0.043555	0.043187	0.041604	0.140162	0.958996	0.12751773
300	11590	13708	13716	0.0965076	0.009395	0	0.035287	0.0391521	1.223709	0.0490786
100	12448	13468	13382	0.029623	0	0	0.009874	0.063871	2.913834	0.02144964
33.3333	12616	11348	12444	0.016526	0.115529	0.029935	0.053986	0.0539965	3.459838	0.013796309
11.1111	12828	12828	12828	0	0	0	0	0	4.459838	0
Control	Cell	12828		Blank	0		Dilution	3	IC50	FALSE

Third batch: (Infergen)

Concentration ( $\times 10^{10}$ /ml)	Inhibition effect to HBsAg						Average inhibiti n rate	Accumulatio n	1- Accumulatio n	Accumulate d inhibition rate
	First well	Second well	Third well	First well	Second well	Third well				
900	7240	5642	6158	0.064599	0.1418	0.20439	0.136951	0.217399	0.862049	0.20131173
300	11072	8796	6902	0	0	0.10826	0.03609	0.0804479	1.82596	0.04217656
100	7015	9746	7852	0.09354	0	0.02428	0.039276	0.044358	2.787663	0.01566301
33.33333	7622	9866	8676	0.015245	0	0	0.005082	0.0050818	3.782601	0.06134167
11.11111	7740	7740	7740	0	0	0	0	0	4.782601	0
Control	Cell	7740		Blank	0		Dilution	3	IC50	FALSE
Concentration ( $\times 10^{10}$ /ml)	Inhibition effect to HBsAg						Average inhibiti n rate	Accumulatio n	1- Accumulatio n	Accumulate d inhibition rate
	First well	Second well	Third well	First well	Second well	Third well				
900	11048	11856	11902	0.04775	0	0	0.015917	0.015917	0.984083	0.01591679
300	13454	12896	11798	0	0	0	0	0	1.984083	0
100	12846	13160	12546	0	0	0	0	0	2.984083	0
33.33333	12680	12458	12360	0	0	0	0	0	3.984083	0
11.11111	11602	11602	11602	0	0	0	0	0	4.984083	0
Control	Cell	11602		Blank	0		Dilution	3	IC50	FALSE

HBsAg: Average IC50: 0 SD: 0

HBsAg: Average IC50: 0 SD: 0

Claims

1. A recombinant super-compound interferon (rSIFN-co) with changed 3-dimensional structure and improved efficacy which can inhibit the DNA duplication and secretion of HBsAg and HBeAg of HBV.
2. The interferon of claim 1, wherein the 3-dimensional change was the result of changes of its production techniques, and efficacy gains not seen in interferon described in U.S. Patent Nos. 4,696,623 and 4,897,471.
3. A super-compound interferon of claim 1 or claim 2, wherein it has its unique secondary and tertiary structure which elicit its special efficacies.
4. A super-compound interferon of claim 1 or claim 2, produced by a highly efficient express system which is constructed with a special promoter.
5. The super-compound Interferon of claim 4, wherein the promoter is P<sub>BAD</sub>.
6. The super-compound interferon of claim 4, wherein its gene is artificially synthesized cDNA, adjusted according to codon preference of *E. Coli*.
7. A process for production of recombinant super-compound interferon recited in claim 1 or 2.
8. The process for production of claim 7, comprising extraction of super-compound interferon from fermentation broth, collection of inclusion body, denaturation and renaturation of the harvested protein.
9. The process of claim 7, wherein the process maintains the high efficacy even when the super-compound interferon is used with an agent and in a particular concentration.
10. The process of claim 7, comprising separation and purification of the super-compound Interferon.
11. The process of claim 7, comprising lyophilization of purified super-compound interferon.
12. The process of claim 7, comprising production of liquid injection of super-compound interferon.
13. Uses of super-compound interferon in preparing medicines for inhibition of HBV-DNA, HBsAg and HBeAg, wherein the virus diseases comprising hepatitis A, hepatitis B, hepatitis C, other types of hepatitis, infections of viruses such as: Epstein-Barr virus, HIV, herpes viruses (Epstein-Barr virus, Cytomegalovirus, herpes simplex viruses), papovaviruses, poxviruses, plomaviruses, adenoviruses, rhinoviruses, human T cell leukaemia viruses I, or human T cell leukaemia viruses II.
14. Uses of claim 1 and 2, wherein the super-compound Interferon selected for Interferon is  $\alpha$ ,  $\beta$ ,  $\gamma$  such as, IFN-1a, IFN-2b or other mutants.
15. Uses of claim 13, wherein super-compound Interferon was administered via oral, vein injection, muscle injection, subcutaneous injection, nasal, or mucosal administration.
16. Uses of claim 13, wherein super-compound Interferon was administered following the protocol as follows: injection 9  $\mu$ g or 15  $\mu$ g per day, 3 times a week, total 24 weeks.



Figure 1

```

5'          11          21          31          41          51
+1 M C D L P Q T H S L G N R R A L I L L A
1 ATGTGTGATT TACCTCAAAC TCATTCTCTT GGTACCGTC GCGCTCTGAT TCTGCTGGCA
TACACACTAA ATGGAGTTTG AGTAAGAGAA CCATTGGCAG CGCGAGACTA AGACGACCGT

5'          71          81          91          1          11
+1 Q M R R I S P F S C L K D R H D F G F F
61 CAGATGCGTC GTATTTCCTT GTTAGCTGC CTGAAAGACC GTCACGACTT CGGCTTTCGG
GTCTACGCAG CATAAAGGGG CAATCGACG GACTTTCTCG CAGTGTCTGAA GCCGAAAGGC

5'          31          41          51          61          71
+1 Q E E F D G N Q F Q K A Q A I S V L H E
121 CAAGAAGAGT TCGATGGCAA CCAATTCCAG AAAGCTCAGG CAATCTCTGT ACTGCACGAA
GTCTTCTCA AGCTACCGTT GGTAAAGTC TTTGAGTCC GTTAGAGACA TGACGTGCTT

5'          91          1          11          21          31
+1 M I Q Q T F N L F S T K D S S A A W D E
181 ATGATCCAAC AGACCTTCAA CCGTGTTCCT ACTAAAGACA GCTCTGCTGC TTGGGACGAA
TACTAGGTTG TCTGGAAGTT GGACAAAGG TGATTCTGT CGAGACGACG AACCTGCTT

5'          51          61          71          81          91
+1 S L L E K F Y T E L Y Q Q L N D L E A C
241 AGCTTGCTGG AGAAGTTCTA CACTGAACGT TATCAGCAGC TGAACGACCT GGAAGCATGC
TCGAACGACC TCTTCAAGAT GTGACTTGAC ATAGTCGTG ACTTGCTGGA CTTTGTGATG

5'          11          21          31          41          51
+1 V I Q E V G V R E T P L M N V D S I L A
301 GTAATCCAGG AAGTTGGTGT AGAAGAGACT CCGCTGATGA ACGTCGACTT TATTCTGGCA
CATTAGGTCC TTCAACCACA TCTTCTCTGA GCGACTACT TGCAGCTGAG ATAAGACCGT

```

## INTERNATIONAL SEARCH REPORT

International application No.

PCT/CN02/00128

## A. CLASSIFICATION OF SUBJECT MATTER

IPC: A61K38/21, A61P1/16, A61P31/12, C12N15/20, C12N15/63, C12N15/70

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC: A61K38/21, A61P1/16, A61P31/12, C12N15/20, C12N15/63, C12N15/70

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Chinese Patents, Chinese Scientific and Technical Journals

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPOQUE, BA, MEDLINE

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO9321229(AMGEN INC et al), 28. Oct. 1993 see abstract	1-16
A	WO8304053(AMGEN INC et al), 24. Nov. 1983 see abstract	1-16

☐ Further documents are listed in the continuation of Box C. ☒ See patent family annex.

## \* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"J" document which may throw doubts on priority claim (5) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&amp;" document member of the same patent family

Date of the actual completion of the international search  
23. July. 2002(23. 07. 02)

Date of mailing of the international search report

08 AUG 2002 (08. 08. 02)

Name and mailing address of the ISA/CN  
6 Xitucheng Rd., Jinan Bridge, Haidian District,  
100088 Beijing, China  
Facsimile No. 86-10-62019451

Authorized officer

SUN, Guangxiu

Telephone No. 86-10-62093884



Form PCT/ISA/210 (second sheet) (July 1998)

**INTERNATIONAL SEARCH REPORT**  
 Information on patent family members

International application No.

PCT/CN02/00128

Patent document cited in search report	Publication date	Patent family members	Publication date
WO 9321229	1993-10-28	US5372808	1994-12-13
		EP0641359	1995-03-08
		CN1081909	1994-02-16
		JP7505894T	1995-06-29
WO 8304053	1983-11-24	EP0108128	1984-05-16
		JP59501097T	1984-06-28
		US4897471	1990-01-30

Form PCT/ISA/210 (patent family annex) (July 1998)